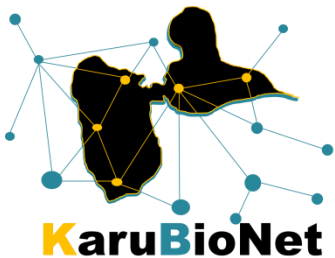


# Bacterial sequencing on ONT MinION

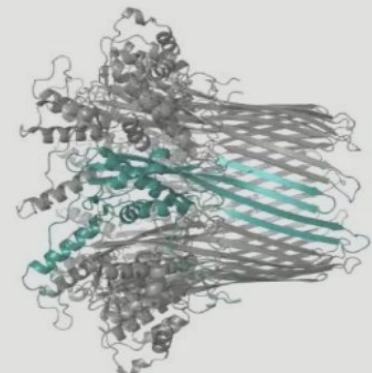
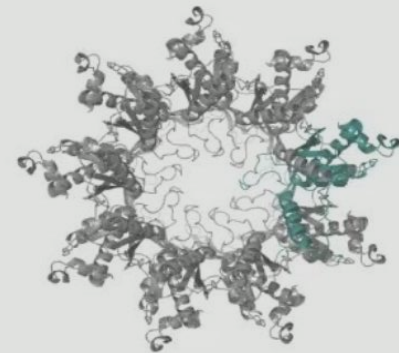
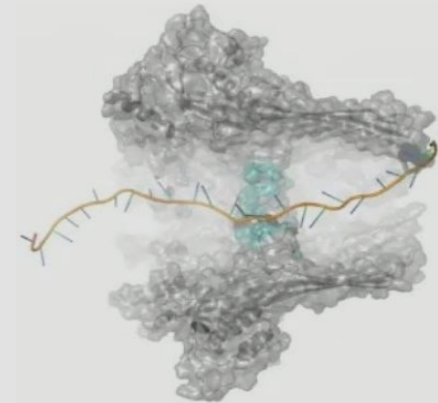
---

Yann Reynaud

KaruBioNet meeting 13/11/2019



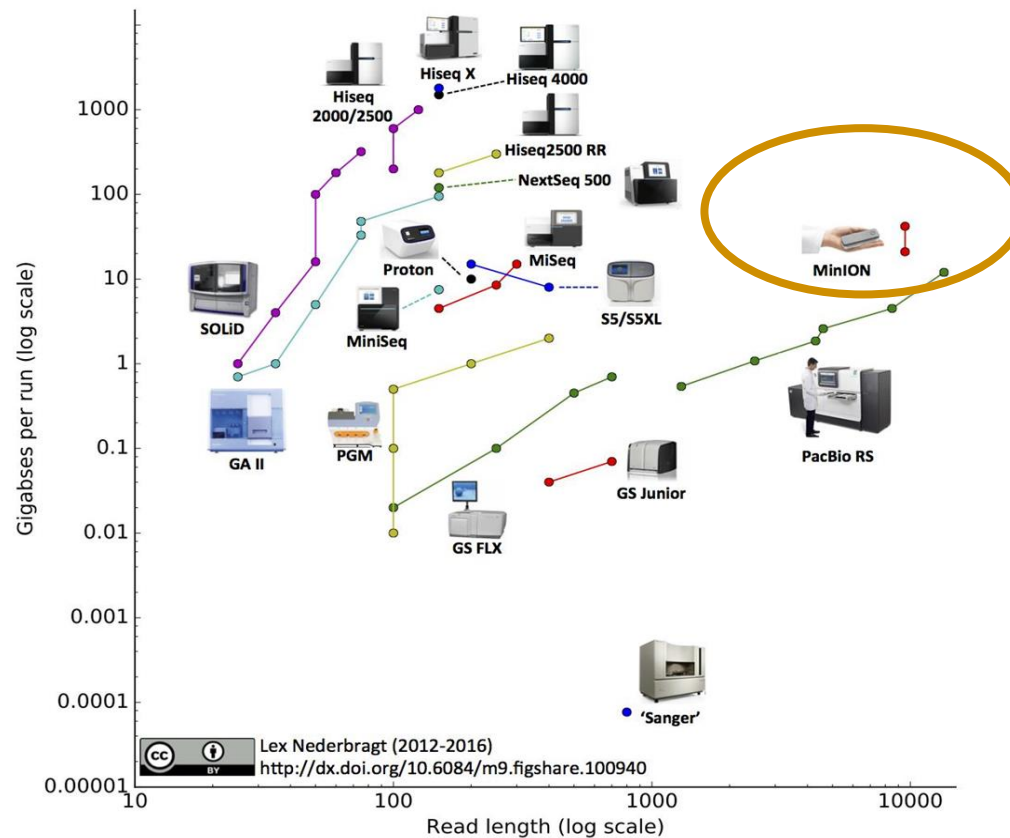
Transmission Réservoirs &  
Diversité des Pathogènes



# Context

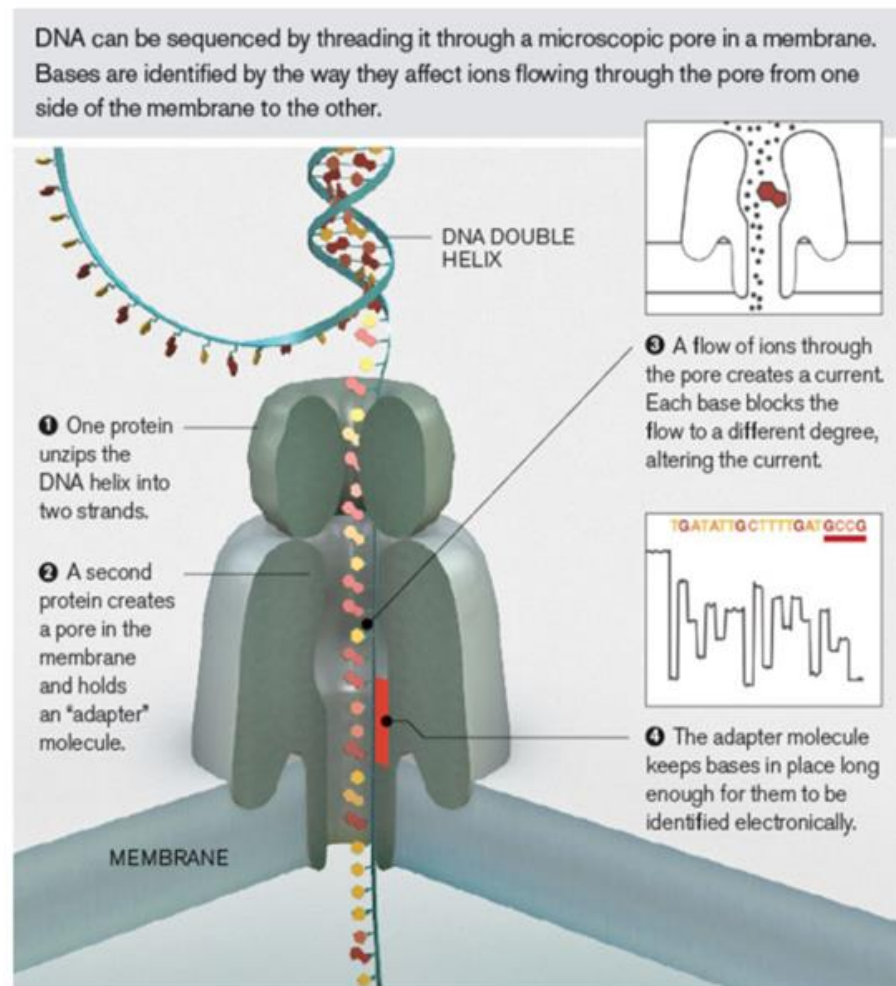
## ●●● 3rd generation sequencing

Single-molecule direct sequencing (“third-gen”) vs. sequencing by synthesis (e.g. Illumina)



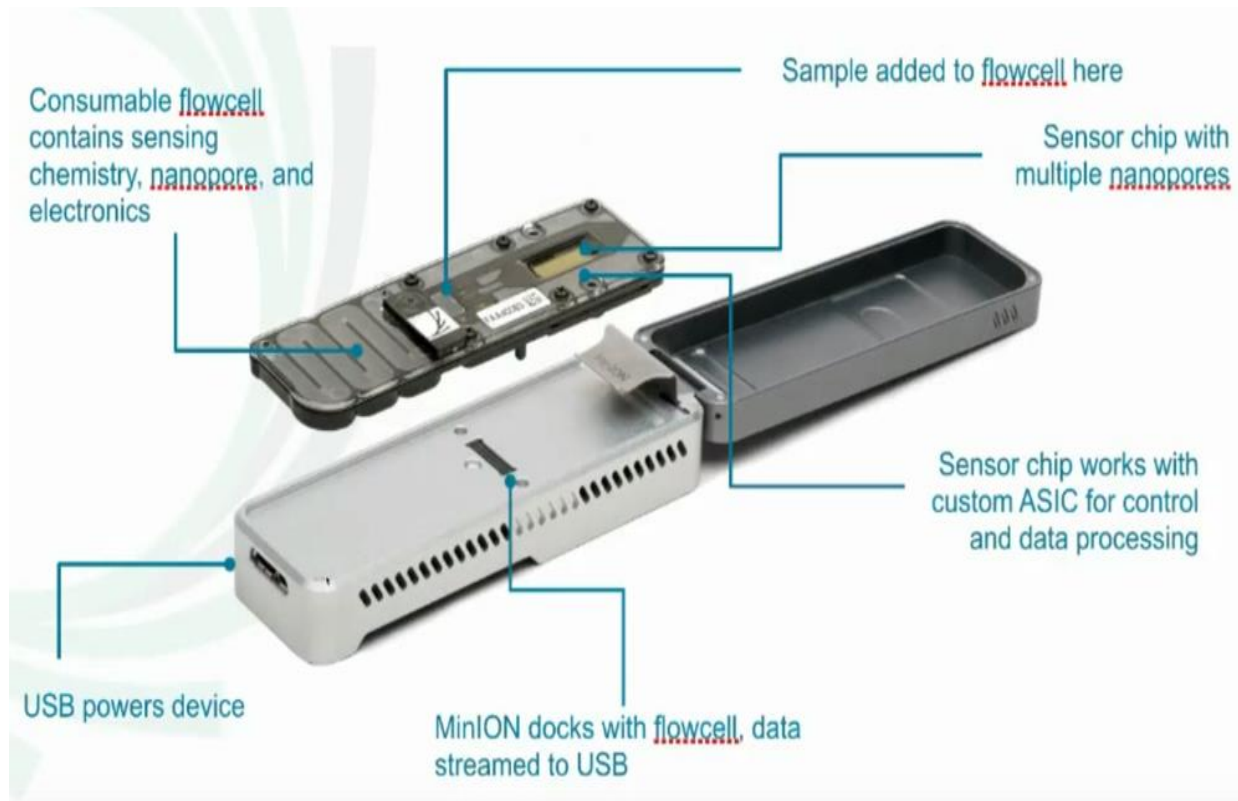
# Context

## ●●● How it works ?



# Context

## ●●● Anatomy



# Context

## ●●● Fast evolution



# Context

## ●●● Fast evolution

	Flongle	MinION Mk1B	MinION Mk1C	GridION Mk1	PromethION 24
Number of channels per flow cell	126	512	512	512	3000
Number of flow cells per device	1	1	1	5	24
Price per flow cell	\$90	\$900 - \$475	\$900 - \$475	\$900 - \$475	\$2000 - \$625
Run time	1 min - 16 hours	1 min - 48 hours	1 min - 48 hours	1 min - 48 hours	1 min - 72 hours
DNA sequencing yield per flow cell (typical in field - best in field)	1 - 2 Gb	15 - 30 Gb	15 - 30 Gb	15 - 30 Gb	100 - 180 Gb
DNA sequencing yield per device (typical in field - best in field)	1 - 2 Gb	15 - 30 Gb	15 - 30 Gb	75 - 150 Gb	2.4 - 4.3 Tb
Price per Gb, DNA sequencing		@15 - 30 Gb	@15 - 30 Gb	@75 - 150 Gb	@2.4 - 4.3 Tb
	@1 - 2 Gb \$90 per flow cell: 90 - 45	\$900 per flow cell: \$60 - 30 \$790 per flow cell: \$53 - 26 \$675 per flow cell: \$45 - 23 \$500 per flow cell: \$33 - 16 \$475 per flow cell: \$31 - 15	\$900 per flow cell: \$60 - 30 \$790 per flow cell: \$53 - 26 \$675 per flow cell: \$45 - 23 \$500 per flow cell: \$33 - 16 \$475 per flow cell: \$31 - 15	\$900 per flow cell: \$60 - 30 \$790 per flow cell: \$53 - 26 \$675 per flow cell: \$45 - 23 \$500 per flow cell: \$33 - 16 \$475 per flow cell: \$31 - 15	\$2000 per flow cell: \$20 - 11 \$1700 per flow cell: \$18 - 9 \$1475 per flow cell: \$15 - 8 \$1225 per flow cell: \$12 - 7 \$975 per flow cell: \$10 - 5 \$625 per flow cell: \$6 - 3



# Context

## ●●● Steps

New Technology + Startup Company = putatives Troubleshootings !

1. **Purchasing**
2. **Computer**
3. **Software**
4. **DNA extractions**
5. **Library prep**
6. **Sequencing**
7. **Data analysis**
8. **Data storage**
9. **\$ per sample?**



# Steps

## ●●● 1. Purchasing

You need to buy a starter pack before being part of the “Community” ~ 1000 euros

→ 1 minION + 2 flowcells





# Steps

## ●●● 2. Computer

Check requirements otherwise no SAV!

### Minimum host computer specification

This example will allow a user to run a MinION with real-time local basecalling. Given the amount of data potentially generated, the basecalling may fall behind and need to be completed at the end of an experiment.

Component	Configuration		
Operating System	Windows	Mac	Linux
	7, 8, 10 (64 bit)	Yosemite	Ubuntu 14.4*
		El Capitan	16.4
		Sierra (64 bit)	
Memory/RAM:	16GB RAM		
CPU	i7 or Xeon**		
Storage	1TB SSD		
Ports	USB3***		
<i>* Linux products are offered under limited support and may take the team longer to respond to queries</i>			
<i>** Users need to verify their i7 is a four core model or better.</i>			
<i>*** The MinION device is CE marked using USB3. If a user wished to use USB-C they may but this invalidates the CE marking</i>			



# Steps

## ●●● 3. Softwares



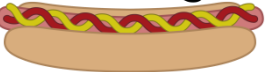
**MinKNOW**, the operating system

- Data acquisition
- Real time analyses & feedback
- Local basecalling
- Output .fast5 or .fastq files



**Guppy** contains the ONT basecalling algorithms  
(NB other basecaller possible)

Filtlong



**Filtlong**

Filer short reads, and decrease randomly reads if too many data generated



# Steps

## ●●● 3. Softwares



### EPI2ME: several workflows

**> CONNECTION TEST**

FASTQ CONNECTION TEST 3.10.4

Test your EPI2ME configuration

5 €

**ANTIMICROBIAL RESISTANCE**

FASTQ ANTIMICROBIAL RESISTANCE 3.3.2

Fastq Antimicrobial Resistance with filter

40 €

**INDEX REFERENCE**

FASTA REFERENCE UPLOAD 3.2.2

Index a reference fasta file for use in custom alignments

10 €

**STRUCTURAL VARIATION**

FASTQ SV CALLER FOR HUMAN 0.0.2

40 €

**HUMAN ALIGNMENT**

FASTQ HUMAN ALIGNMENT GRCh38 3.2.2

Align against Human GRCh38 reference

60 €

**BARCODING**

FASTQ BARCODING 3.10.4

10 €

**16S**

FASTQ 16S 3.2.2

Taxonomic assignment for 16S amplicons

30 €

**FASTQ CONTROL EXPERIMENT**

3.2.2

See how your experiment performed

5 €

**CUSTOM REFERENCE ALIGNMENT**

FASTQ CUSTOM ALIGNMENT 3.2.2

Custom reference alignment

40 €

**CALIBRATION RNA**

FASTQ RNA CONTROL EXPERIMENT 3.2.2

See how your experiment performed

10 €

Allow adapter  
-barcode  
removal + QC

**EXOME**

FASTQ HUMAN EXOME 3.2.2

Align against Human exome

40 €

**What's in my pot?**

FASTQ WIMP 3.2.2

20 €



# Steps

## ●●● 4. DNA extractions



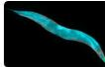
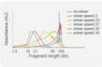


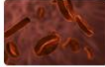


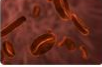










### Key step!

- 1  $\mu\text{g}$  needed quantified Qubit
- OD 260/280 1,8
- OD 260/230 2-2,2
- Check integrity on electrophoresis

Some protocols are proposed

But often you have to optimize your own protocol

Ex: hypervirulent klebsiella cultured with salicilate to remove capsule

Oxford Nanopore-developed methods	Community-developed methods
 <a href="#">Agarose plug</a> View this method	 <a href="#">Avian samples</a> View this method
 <a href="#">C. elegans DNA</a> View this method	 <a href="#">DNA fragmentation</a> View this method
 <a href="#">Drosophila DNA</a> View this method	 <a href="#">FFPE cell line DNA</a> View this method
 <a href="#">Globin mRNA depletion</a> View this method	 <a href="#">Gram-negative bacterial DNA</a> View this method
 <a href="#">Gram-positive bacterial DNA</a> View this method	 <a href="#">Human blood RNA</a> View this method
 <a href="#">Human cell line DNA</a> View this method	 <a href="#">Human cell line RNA</a> View this method
 <a href="#">Human saliva DNA</a> View this method	 <a href="#">Mammalian samples</a> View this method
 <a href="#">mRNA enrichment</a> View this method	 <a href="#">Plant leaf DNA</a> View this method
 <a href="#">Reptilian samples</a> View this method	 <a href="#">Soil DNA</a> View this method
 <a href="#">Size selection</a> View this method	 <a href="#">Stool pellets DNA</a> View this method



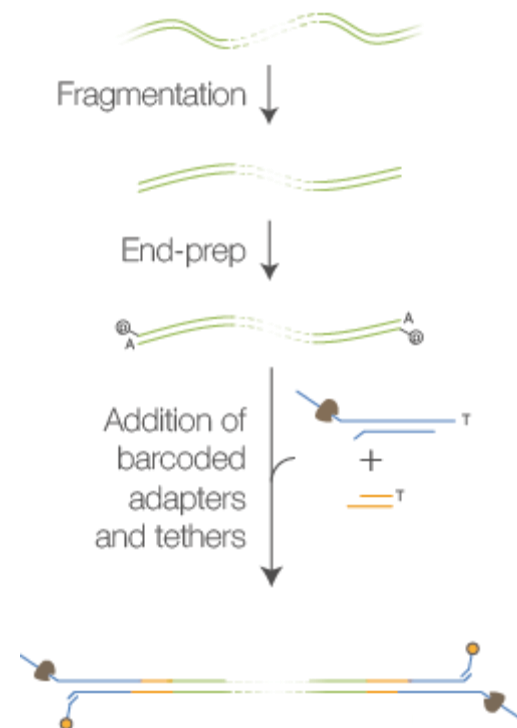
# Steps

## ●●● 5. Library prep

Lot of protocols possible:

- we multiplexed n=7 bacterial samples of *E. coli* and *Klebsiella pneumoniae* (24 possible)
- Fragmentation optional
- Kits ONT
  - Native barcoding Expansion 1-12
  - Ligation sequencing kit
  - Flow cell priming kit
- Additional kits need to be ordered:
  - NEBNext FFPE Repair mix
  - NEBNext End repair d-A tailing
  - NEB Blunt/TA ligase master mix
  - NEBNext quick ligation module
  - LoBind tubes

b) PCR-free barcoding



# Steps

## ●●● 5. Library prep

Basecalled	N°Souche	DO260/280nmGenovaNano	DO260/230nmGenovaNano	Concentration[ng/μl]Qubit	concentration_after_barcode	volume_kept_for_pooling DNA_kept(ng)	ngtotal
BC1	GESC241	1,8	1,4	65,6	46.6	7	326
BC2	GESC242	1,8	1,5	111	38.6	7	270
BC3	GESC249	1,9	1,3	46,4	43.4	7	303
BC4	GESC272	1,9	1,5	63,9	44.8	7	313
BC5	GESC324	1,7	1,2	49,5	55.4	9	498
BC6	SF07	1,3	2,7	3,7	7.74	14	108
BC10	KPB6	1,7	0,5	11,1	14	14	196

sample	DO260/280nmGenovaNa no	DO260/230nmGenovaNa no	Concentration[ng/μl]Qubit
pooled samples	1,789	1,432	60
volume total loaded: 12 μl. Total DNA 720 ng			



# Steps

## ●●● 6. Sequencing & 7. Data analyses

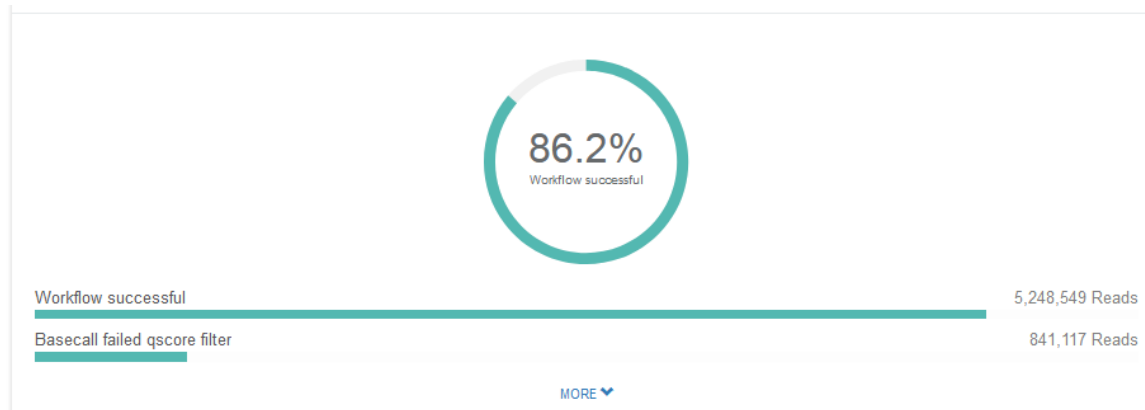
- First check number of pores available (bad if <800): here 1270/2000
- Time determine sequencing depth: can be stopped whenever needed.  
Here 48h
- Only 30% basecalled when the run was stopped
- Basecalling finished on guppy after recovering raw files: very long on classical CPU. So run on UA cluster GPU... 1 day against 1 month!!!



# Steps

## ●●● 7. Data analyses

- EPI2ME for QC, demultiplexing



☰ Reads per Barcode ID

ID ▾	Read Count	
BC01	799,841	EXCLUDE
BC02	482,411	EXCLUDE
BC03	983,965	EXCLUDE
BC04	894,275	EXCLUDE
BC05	1,277,528	EXCLUDE
BC06	545,093	EXCLUDE
BC07	73	INCLUDE
BC08	24	INCLUDE
BC09	81	INCLUDE
BC10	22,661	EXCLUDE





# Steps

## ●●● 7. Data analyses

- EPI2ME for QC, demultiplexing

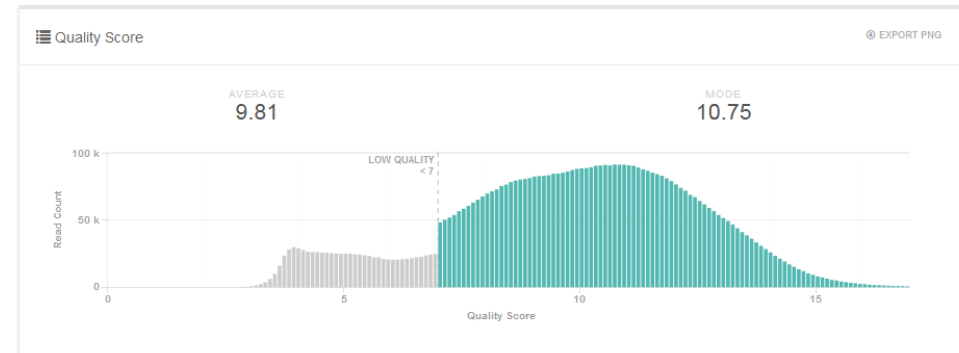
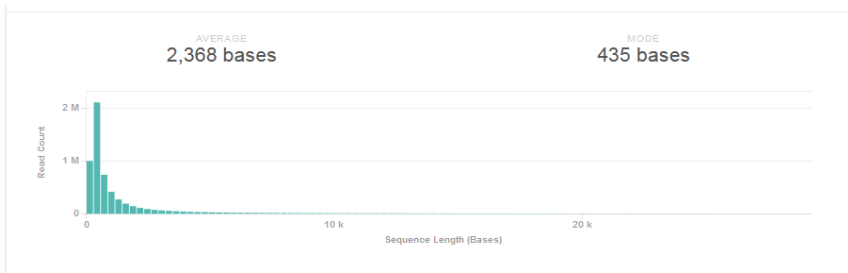
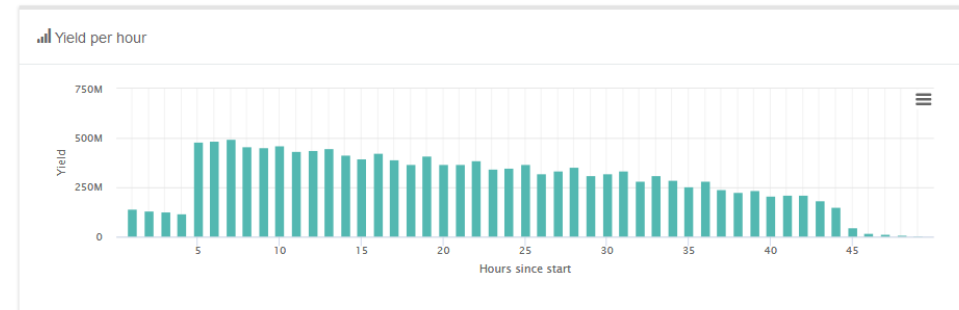
Failed for one sample

Relatively small fragments

But great depth

Good quality score

Basecalled	N°Souche	reads_count	~coverage(5Mgenome)
BC1	GESC241	799841	440X
BC2	GESC242	482411	440X
BC3	GESC249	983965	340X
BC4	GESC272	894275	420X
BC5	GESC324	1277528	560X
BC6	SF07	545093	260X
BC10	KPB6	22661	7.92X



# Steps

## ●●● 7. Data analyses

### Unicycler:

Hybrid assembling between illumina and ONT:

- 1/ gaps in short read assembly graph are bridged with long read contigs ;
- 2/ contigs are circularized, overlaps removed and reorientated to dnaA/repA (if possible)
- 3/ final contigs are polished with short reads data

Handles plasmid-rich genomes

Low misassembly rates

Deal with repetitive genomes

Produce assembly graph for Bandage



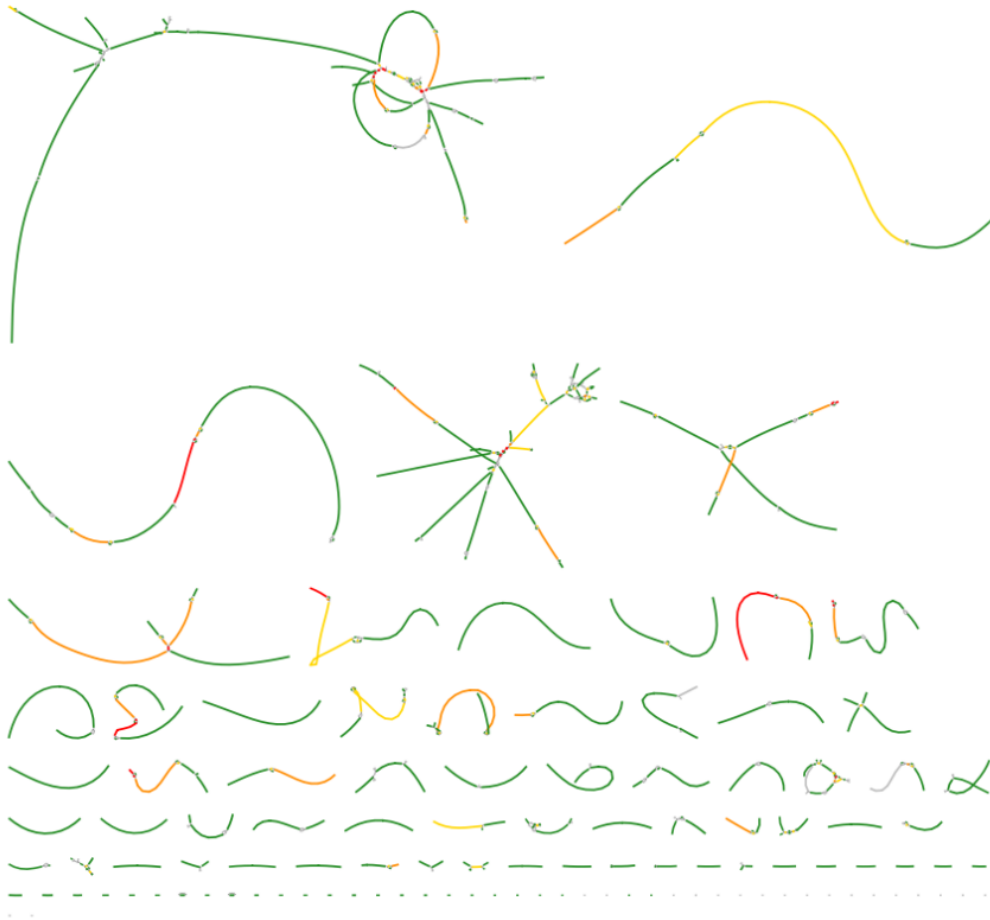
Bandage



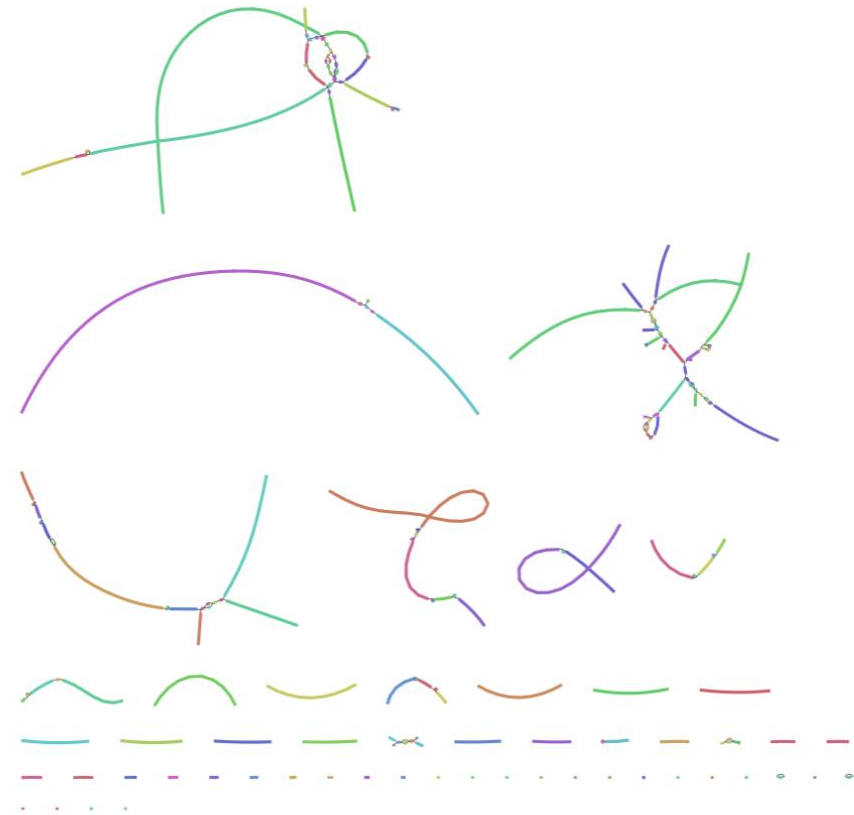
# Steps

## ●●● 7. Data analyses

Illumina



Hybrid Illumina/ONT



# Steps

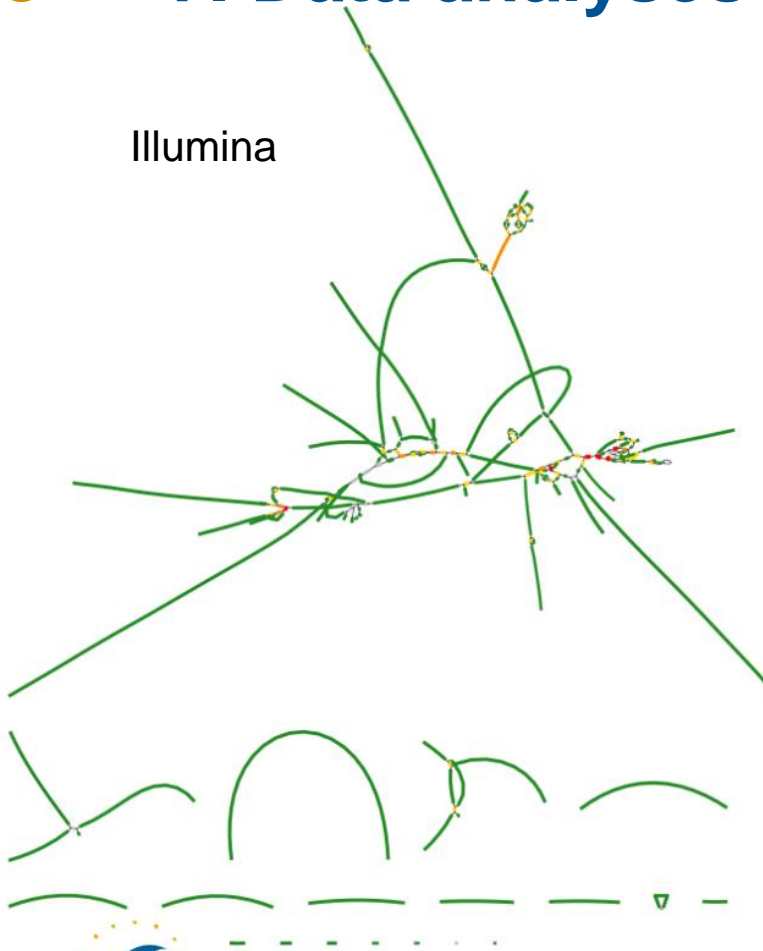
## ●●● 7. Data analyses



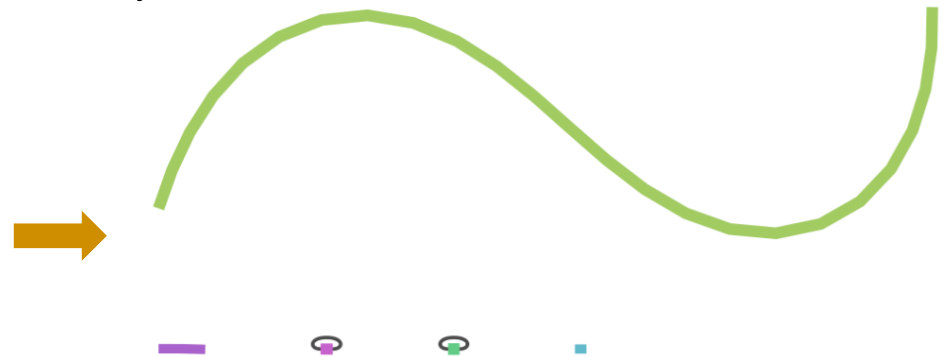
# Steps

## ●●● 7. Data analyses

Illumina



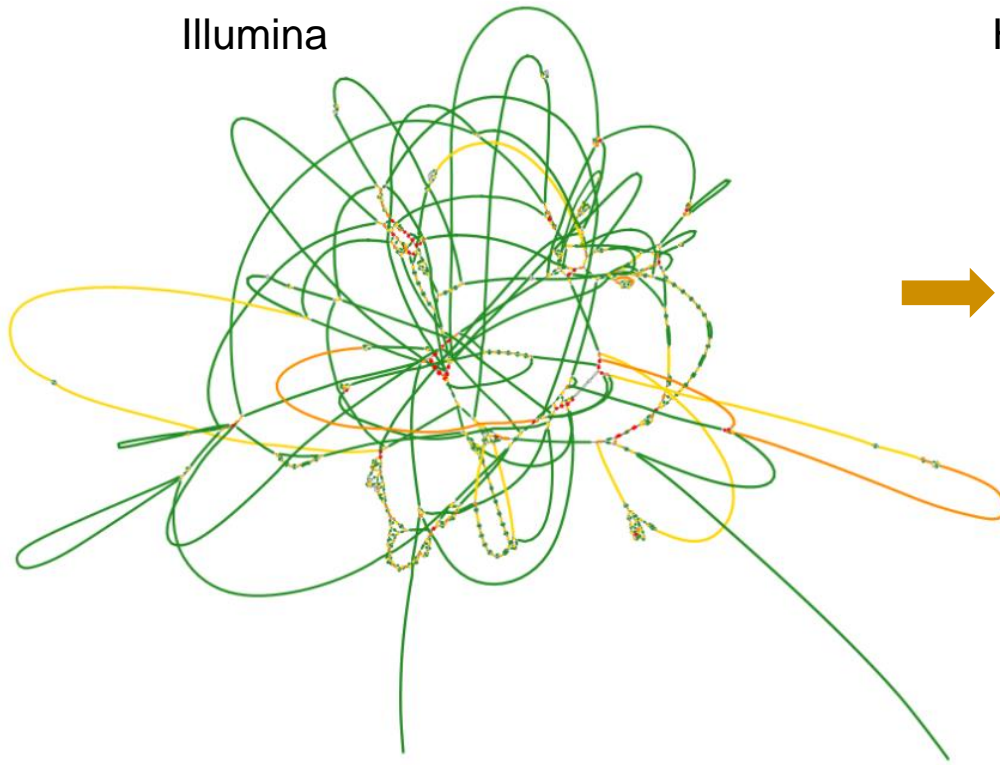
Hybrid ONT/Illumina



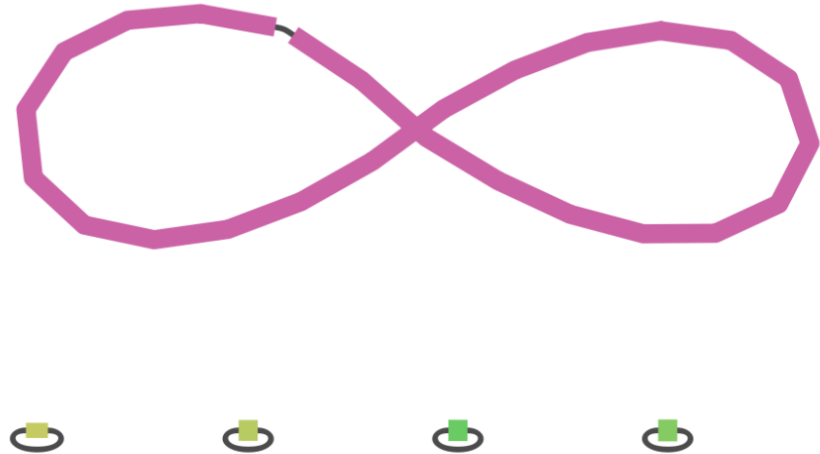
# Steps

## ●●● 7. Data analyses

Illumina

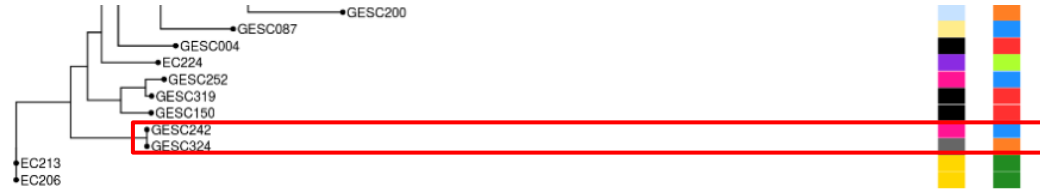


Hybrid ONT/Illumina PERFECT

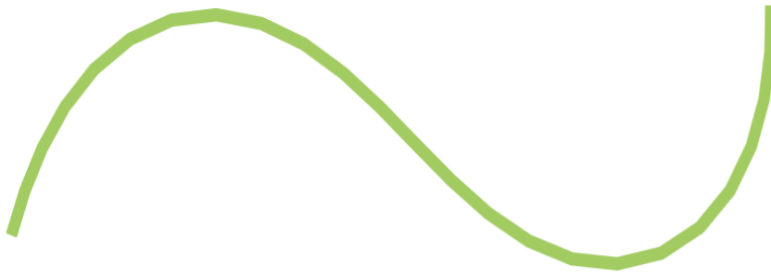


# Steps

## ●●● 7. Data analyses



GESC242 (oiseau, STEP)



#GESC242							
Component	Segments	Links	Length	N50	Longest segment		
total	5	2	5,040,336	4,745,166	4,745,166		
1	1	0	4,745,166	4,745,166	4,745,166	incomplete	
2	1	0	204,105	204,105	204,105	incomplete	
3	1	1	53,644	53,644	53,644	complete	
4	1	1	34,653	34,653	34,653	complete	
5	1	0	2,768	2,768	2,768	incomplete	

GESC324 (cafard, égout CHU)



# GESC324							
Component	Segments	Links	Length	N50	Longest segment		Status
total	4	2	5,133,535	4,012,746	4,012,746		
1	1	0	4,012,746	4,012,746	4,012,746	incomplete	
2	1	1	803,514	803,514	803,514	complete	
3	1	1	309,211	309,211	309,211	complete	
4	1	0	8,064	8,064	8,064	incomplete	



# Steps

## ●●● 7. Data analyses

### Resfinder database

#### GESC242



blaCTX-M-15  
blaOXA-1  
aac(6')-Ib-cr  
tet(A)  
aac(3)-IIa  
aac(6')-Ib-cr  
dfrA14  
catB3

blaTEM-1B  
qnrB1  
qnrS1  
aph(3'')-Ib  
aph(6)-Id  
dfrA14

204,105    53,644    IncN

#### GESC324



probably chromosomal?  
Chro 4M  
Contig 2 800 kb

blaCTX-M-15  
blaOXA-1  
blaTEM-1B  
aac(6')-Ib-cr  
qnrB1  
tet(A)  
aac(3)-IIa  
aac(6')-Ib-cr  
aph(3'')-Ib  
aph(6)-Id  
dfrA14  
catB3  
sul2

IncH12  
309,211

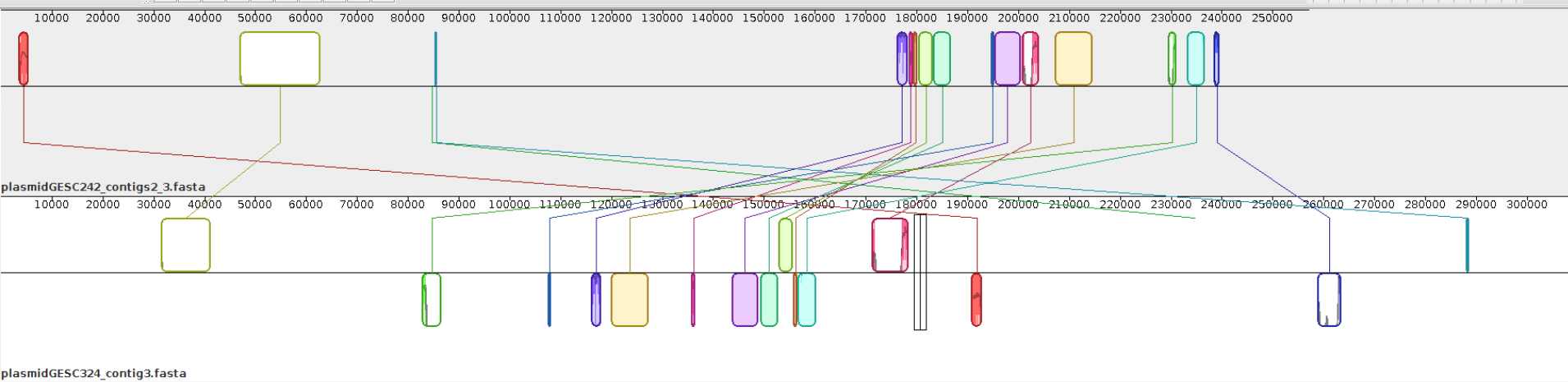




# Steps

## ●●● 7. Data analyses

Mauve syntheny: seems to be different plasmids sharing close ATBR profiles  
OR  
Assembly with error due to repetitive elements unresolved by ONT (to short read ~2500 kb)



# Steps

## ●●● 8. Data storage

~ 5 Go / ONT .fastq file (for bacteria)

Take care to remove all files when finished

If you perform live basecalling and if you have less than 1To available, MinKNOW might crash!



# Steps

## ●●● 9. Cost

Computer 1To SSD, i7, 16 Go ram= 1400 euros

Barcoded \*12 bacteria <100 euros/strain

Putatively 50 euros if barcoded \*24! (~ Illumina)



# Conclusion

- gDNA extraction needs to be optimized
  - Increase size of fragments to resolve repetitive elements
  - Quality of DNA should be improved 260/230
  - Protocol adjusted for Kp hypervirulent
- Basecalling done on Guppy at UA instead of live basecalling
- Barcoding \*24 should be tested
- PFGE to check that we sequence correct size of plasmids?

